

DATASHEET: LYOPHILIZED TUBULIN (>99%)

Catalog Number: 142001

Source: Bovine Brain

Store Desiccated at 4°C

Background:

Tubulin, a highly conserved cytoskeletal protein, is required for several essential eukaryotic processes including intracellular transport, intercellular signaling, extracellular sensing, cell migration, and cell division. Tubulin (110 kDa) is a heterodimer of α - and β -tubulin (each 55 kDa), and polymerizes into higher order filaments termed microtubules (MTs). MTs measure 25 nm in diameter and have a persistence length of \sim 2 μ m, incorporating \sim 1650 tubulin subunits per 1 μ m. Given the asymmetry of tubulin dimers, MTs have inherent polarity with distinct “+” (β -tubulin exposed) and “-” (α -tubulin exposed) ends. Another critical feature of MTs is their dynamic instability, a consequence of the GTPase activity of tubulin. This property confers force-generating capabilities to MTs that are critical for cell division. For this reason, tubulin is a powerful target for the therapeutic intervention of neoplastic diseases such as cancers.

Material:

Lyophilized Tubulin is isolated by selective precipitation from bovine brain homogenate by an adaptation of the method of Andreu (2007) and lyophilized by an adaptation of the method of Dráberová *et al.* (2010). The resulting product is >99% pure (Figure 1) and polymerization competent (Figure 2). Lyophilized Tubulin is supplied as a white powder with a tubulin-to-powder weight ratio of 1:2. When reconstituted with Milli-Q water to 40 mg/ml, the buffer conditions are 10 mM Sodium Phosphate, 0.5 mM MgCl₂, 0.1 mM GTP, and 0.25 M Trehalose, pH 7.0.

Storage and Handling:

Store Lyophilized Tubulin desiccated at 4°C. The product is stable under these conditions for 1 year. Reconstitute Lyophilized Tubulin by resuspending in ice-cold Milli-Q water to 40 mg/ml (note that 1 mg tubulin = 3 mg powder). Add EGTA and MgCl₂ to 1 mM each, and clarify the solution at 14,000 rpm for 1 minute at 4°C. Measure the protein concentration and dilute to 20 mg/ml with Tubulin Phosphate Buffer (Cat. No. 142002; 10 mM Sodium Phosphate, 1 mM EGTA, and 1 mM MgCl₂, pH 7.0). Reconstituted Lyophilized Tubulin can be buffer-exchanged or cycled as desired, as well as frozen in liquid Nitrogen and stored at -80°C. Avoid repeated freeze-thaw cycles.

Activity:

When supplemented with guanosine (GTP or GMPCPP) and warmed to 37°C, Lyophilized Tubulin will polymerize into MTs when above its critical concentration. The recommended tubulin concentration for ensuring polymerization is 2 mg/ml.

Uses:

Lyophilized Tubulin is supplied for use in cell-free experimental systems including:

- structural analysis by X-ray crystallography and electron microscopy
- drug discovery by high-throughput screening
- *in vitro* biochemical and biophysical approaches

Polymerization Protocol:

Dilute reconstituted Lyophilized Tubulin to 2 mg/ml with Tubulin Phosphate Buffer (Cat. No. 142002; 10 mM Sodium Phosphate, 1 mM EGTA, and 1 mM MgCl₂, pH 7.0) and supplement with 1 mM each DTT and guanosine (GTP or GMPCPP). Incubate on ice for 5 minutes, then transfer to a 37° C water bath for 1 hour. If polymerized with GMPCPP or protected with Taxol, the resulting MTs will be stable at room temperature for several days. Do not place polymerized MTs on ice.

Technical Notes:

- store desiccated at 4°C
- flash-freeze in experimental-sized aliquots upon reconstitution
- regard tubulin concentration, temperature, and guanosine addition when polymerizing
- do not place polymerized MTs on ice

Figure 1: Lyophilized Tubulin is >99% pure. Coomassie G250-stained protein gel of Lyophilized Tubulin separated by SDS-PAGE. The tubulin appears as a single species migrating at ~55 kDa. Molecular weight markers (kDa) and loaded protein quantities are indicated.

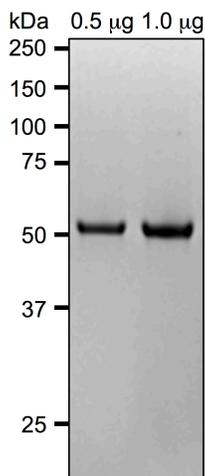
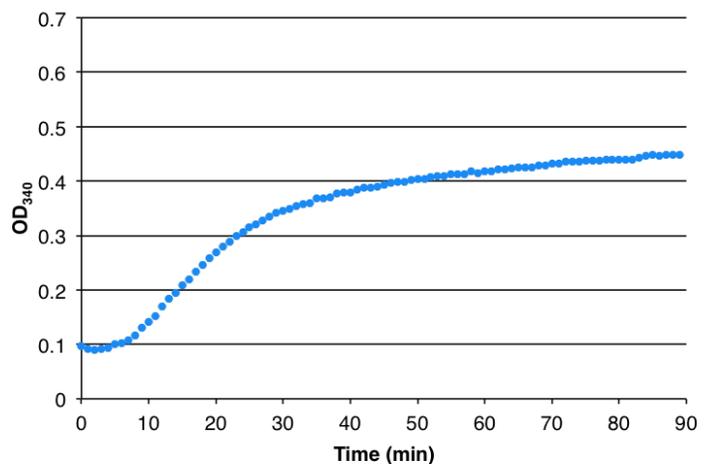


Figure 2: Lyophilized Tubulin is polymerization-competent. Optical Density (340 nm) of Lyophilized Tubulin at 5 mg/ml in Tubulin PEM Buffer (Cat. No. 032002; 80 mM PIPES, 1 mM EGTA, and 1 mM MgCl₂, pH 6.8) supplemented with 1 mM GTP, 1 mM DTT, and 5% Glycerol at 37°C. Distinct nucleation and polymerization phases are evident.



Comparison with Cycled Tubulin™ (Cat. No. 032005):

Specifications	Cycled Tubulin™	Lyophilized Tubulin
Cat. No.	032005	142001
Purity	>99%	>99%
Cycled	Yes	No
Storage Method	Cryopreserved	Lyophilized
Storage Buffer	PIPES	Phosphate
Price	< \$150/mg	< \$105/mg
Shipping Method	FedEx Overnight on Dry Ice	FedEx 2 Day Envelope

References:

1. Allen, C. & Borisy, G. G. Structural polarity and directional growth of microtubules of *Chlamydomonas flagella*. *J. Mol. Biol.* **90**, 381–402 (1974).
2. Andreu, JM. Large scale purification of brain tubulin with the modified Weisenberg procedure. *Methods Mol. Med.* **137**, 17-28 (2007).
3. Caplow, M., Ruhlen, R. L. & Shanks, J. The free energy for hydrolysis of a microtubule-bound nucleotide triphosphate is near zero: all of the free energy for hydrolysis is stored in the microtubule lattice. *J. Cell Biol.* **127**, 779–788 (1994).
4. Carrier, M. F., Hill, T. L. & Chen, Y. Interference of GTP hydrolysis in the mechanism of microtubule assembly: an experimental study. *P Natl Acad Sci Usa* **81**, 771–775 (1984).
5. David-Pfeuty, T., Erickson, H. P. & Pantaloni, D. Guanosinetriphosphatase activity of tubulin associated with microtubule assembly. (1977).
6. Desai, A. & Mitchison, T. J. Microtubule polymerization dynamics. *Annu. Rev. Cell Dev. Biol.* **13**, 83–117 (1997).
7. Dráberová, E., Sulimenko, V., Sulimenko, T., Böhm, K., & Dráber, P. Recovery of tubulin functions after freeze-drying in the presence of trehalose. *Analytical Biochemistry.* **397**, 67–72 (2010).
8. Evans, L., Mitchison, T. & Kirschner, M. Influence of the centrosome on the structure of nucleated microtubules. *J. Cell Biol.* **100**, 1185–1191 (1985).
9. Mandelkow, E. M., Mandelkow, E. & Milligan, R. A. Microtubule dynamics and microtubule caps: a time-resolved cryo-electron microscopy study. *J. Cell Biol.* **114**, 977–991 (1991).
10. Mitchison, T. & Kirschner, M. Dynamic instability of microtubule growth. *Nature* (1984).
11. Nogales, E., Whittaker, M., Milligan, R. A. & Downing, K. H. High-Resolution Model of the Microtubule. *Cell* **96**, 79–88 (1999).
12. Oosawa, F., 1922, Asakura, S. 1927. Thermodynamics of the polymerization of protein. (1975).
13. Walker, R. A. *et al.* Dynamic instability of individual microtubules analyzed by video light microscopy: rate constants and transition frequencies. *J. Cell Biol.* **107**, 1437–1448 (1988).
14. Weisenberg, R. C., Deery, W. J. & Dickinson, P. J. Tubulin-nucleotide interactions during the polymerization and depolymerization of microtubules. *Biochemistry* **15**, 4248–4254 (1976).